The amphibian chytrid fungus, *Batrachochytrium dendrobatidis* (*Bd*) is known to have a wide distribution in North America (http://www.bd-maps.net/), but confirmed infections in areas of the American Mid-South are lacking. We surveyed an ephemeral water body in Wapanocca National Wildlife Refuge (WNWR), Turrell, Arkansas, USA. While other surveys have confirmed the presence of *Bd* in Arkansas via histology (Rothermel et al. 2008), no survey thus far has confirmed infections via the more-quantitative real-time polymerase chain reaction (qPCR) method (Kerby 2012). Our survey was the first to quantitatively assess *Bd* presence on amphibians and their infection intensities in Arkansas.

On 20 May 2013, we surveyed Pecan Ridge ephemeral depression (35.33113°N, 90.221467°W) within WNWR (Fig. 1). With the assistance of the Ecological Techniques Class from Christian Brothers University (CBU), we captured amphibians in this area with dipnets, water seines, and by hand. To sample for *Bd*, animals were swabbed using sterile cotton-tipped applicators (Fisherbrand®, Cat. #23-400-116), approximately 30 times along keratinized tissues (ventral surface and hind limbs) where *Bd* zoospores are concentrated (Marantelli et al. 2004). The swabs were placed in 1.5-mL snap-cap tubes (Fisherbrand®, Cat. # 02-681-272) and stored in a -18°C freezer until qPCR analysis was conducted to quantify *Bd* loads and prevalence. To minimize cross-contaminating animals and ensure that our results were accurate, gloves and equipment that contacted frogs were sterilized with a 10% bleach solution or discarded after leaving each water body (Johnson et al. 2003). Infection status (*Bd*-/+) of all animals was confirmed using qPCR following a method by Kerby et al. (2012). DNA was extracted from cotton swabs using Qiagen DNeasy Blood and Tissue Kit (Cat. #69506), and the extracted DNA was used in the qPCR reaction. Standards for the reaction were obtained from Commonwealth Scientific and Industrial Research Organisation laboratories, Australia, and were the same as those used in Boyle et al. (2004). The standards served as the positive controls (versus internal controls, Kerby et al. 2012) and each plate contained a negative control (which tested negative on all plates) and all samples were run in triplicate. For determination of infection status, swabs were categorized as *Bd*-positive when zoospore equivalents were ≥ 1 (as used by Vredenburg et al. 2010).

We captured and swabbed 46 adult Cricket Frogs (*Acris crepitans*). We detected *Bd* in 42 of 46 (87%) animals. In infected individuals, zoospore loads ranged from ~2 to >200,000 (it must be noted that the highest standard used in our experiment is 100 zoospore equivalents). No frog showed clinical signs of *Bd* infection (e.g., lethargy, skin sloughing), indicating that Cricket Frogs may persist in populations without succumbing to *Bd*-induced mortality (but see Rothermal et al. 2008). Our data is the first to show the presence of *Bd* in Arkansas when confirmed through histology (Rothermel et al. 2008), but confirmed infections in areas of the American Mid-South are lacking. We surveyed an ephemeral water body in Wapanocca National Wildlife Refuge (WNWR), Turrell, Arkansas, USA. While other surveys have confirmed the presence of *Bd* in Arkansas via histology (Rothermel et al. 2008), no survey thus far has confirmed infections via the more-quantitative real-time polymerase chain reaction (qPCR) method (Kerby 2012). Our survey was the first to quantitatively assess *Bd* presence on amphibians and their infection intensities in Arkansas.

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Within North America, *Batrachochytrium dendrobatidis* (*Bd*) infections have primarily been reported in the western and northeastern parts of the United States (http://www.bd-maps.net/, Olson et al. 2013). Infections in other parts of the country (e.g., Tennessee) are currently recorded at a much lower incidence, and so further survey data are needed to inform scientists and managers of *Bd* occurrences and risks to native fauna. Chatfield et al. (2009) confirmed *Bd* infections in the Great Smoky Mountains of Tennessee and Davis et al. (2012) confirmed *Bd* infections in Shelby County, Tennessee. Both of these studies confirmed *Bd* infection status via quantitative real time polymerase chain reaction (qPCR), the accepted standard in the field. However, *Bd* also was detected in Meeman-Shelby State Park, Tennessee (Venesky and Brem 2008), but unlike the other surveys in Tennessee, infection was only confirmed in a single individual via histology. To better understand the distribution of *Bd* in areas near Meeman-Shelby State Park, we surveyed ponds and lowlands adjacent to Meeman-Shelby State Park, specifically within the University of Memphis, Edward J. Meeman Biological Field Station (MBFS, 35.366667°N, 90.016667°W, Fig. 1).

On 8 May and 31 May 2013, we captured adult amphibians for *Bd* sampling at four sites within MBFS using dipnets, water seines, and by hand capture. Captures on 8 May were taken to the laboratory at the University of Memphis, Memphis, Tennessee for other projects and thus were not released back into the wild. This ensured that we did not recapture and resample amphibians. To sample *Bd*, animals were swabbed using sterile cotton-tipped applicators (Fisherbrand, Cat. #23-400-116), approximately 30 times along the ventral surface and hind limbs (areas

**Fig. 1.** County map of eastern Arkansas, USA. Wapanocca National Wildlife Refuge is located in Turrell, Arkansas (star within Crittenden Co.). Specific sampling site is indicated by the star within the inset of Wapanocca National Wildlife Refuge.